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(54) Title: PROCESS FOR CONTROLLING LEPI	DOPTE	RAN PESTS
(57) Abstract		

The subject disclosure concerns Bacillus thuringlensis strains which can be used to control lepidopteran pests. The strains were previously known to control coleopteran pests. The discovery of lepidopteran activity was totally unexpected. These B.t. strains can be formulated using standard lepidopteran formulation procedures. Means of administration are also standard. The genes encoding lepidopteran-active toxins can be isolated from the B.t. isolates and used to transform other microbes or plants for use to control lepidopteran pests.

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DESCRIPTION

PROCESS FOR CONTROLLING LEPIDOPTERAN PESTS

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Cross-Reference to Related Applications

This application is a continuation-in-part of co-pending U.S. application Serial No. 07/758,020, filed September 12, 1991, which is a continuation-in-part of co-pending U.S. application Serial No. 07/642,112, filed January 16, 1991, now abandoned. Serial No. 07/758,020 is also a continuation-in-part of U.S. application Serial No. 07/658,935, filed February 21, 1991.

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Background of the Invention

The soil microbe *Bacillus thuringiensis* (*B.t.*) is a Gram-positive, spore-forming bacterium characterized by parasporal crystalline protein inclusions. These often appear microscopically as distinctively shaped crystals. The proteins are highly toxic to pests and specific in their activity. The toxin genes have been isolated and sequenced, and recombinant DNA-based *B.t.* products produced and approved. In addition, with the use of genetic engineering techniques, new approaches for delivering *B.t.* endotoxins to agricultural environments are under development, including the use of plants genetically engineered with endotoxin genes for insect resistance and the use of stabilized intact microbial cells as *B.t.* endotoxin delivery vehicles (Gaertner, F.H., L. Kim [1988] *TIBTECH* 6:S4-S7). Thus, isolated *B.t.* endotoxin genes are becoming commercially valuable.

Bacillus thuringiensis produces a proteinaceous paraspore or crystal which is toxic upon ingestion by a susceptible insect host. Over the past 30 years, commercial use of B.t. pesticides has been largely restricted to a narrow range of lepidopteran (caterpillar) pests. Preparations of the spores and crystals of B. thuringiensis subsp. kurstaki have been used for many years as commercial insecticides for lepidopteran pests. For example, B. thuringiensis var. kurstaki HD-1 produces a crystal called a delta endotoxin which is toxic to the larvae of a number of lepidopteran insects.

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In recent years, however, investigators have discovered B.t. pesticides with specificities for a much broader range of pests. For example, other species of B.t., namely israelensis and san diego (a.k.a. B.t. tenebrionis, a.k.a. M-7), have been used commercially to control insects of the orders Diptera and Coleoptera, respectively (Gaertner, F.H. [1989] "Cellular Delivery Systems for Insecticidal Proteins: Living and Non-Living Microorganisms," in Controlled Delivery of Crop Protection Agents, R.M. Wilkins, ed., Taylor and Francis, New York and London, 1990, pp. 245-255). See also Couch, T.L. (1980) "Mosquito Pathogenicity of Bacillus thuringiensis var. israelensis," Developments in Industrial Microbiology 22:61-76; Beegle, C.C., (1978) "Use of Entomogenous Bacteria in Agroecosystems," Developments in Industrial Microbiology 20:97-104.



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Krieg, A., A.M. Huger, G.A. Langenbruch, W. Schnetter (1983) Z. ang. Ent. 96:500-508, describe a B.t. isolate named Bacillus thuringiensis var. tenebrionis, which is reportedly active against two beetles in the order Coleoptera. These are the Colorado potato beetle, Leptinotarsa decembrata, and Agelastica alni.

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Recently, many new subspecies of Rt have been identified, and many genes responsible for active &-endotoxin proteins have been isolated (Hofte, H., H.R. Whiteley [1989] Microbiological Reviews 52(2):242-255). Hofte and Whiteley classified 42 Rt crystal protein genes into 14 distinct genes, grouped into 4 major classes based on amino-acid sequence and host range. The classes were Cryl (Lepidoptera-specific), Cryll (Lepidoptera- and Diptera-specific), Cryll (Coleoptera-specific), and CrylV (Diptera-specific). The discovery of strains specifically toxic to protozoan pathogens, animal-parasitic liver flukes (Trematoda), or mites (Acarl) has broadened the potential Rt product spectrum even further. With activities against unique targets, these novel strains retain their very high biological specificity; nontarget organisms remain unaffected. The availability of a large number of diverse Rt toxins may also enable farmers to adopt productures strategies that minimize the risk that Rt-resistant pests will arise.

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The cloning and expression of a Br. crystal protein gene in Escherichia coli has been described in the published literature (Schnepf, H.E., H.R. Whitely [1981] Proc. Natl. Acad. Sci. USA 78:2893-2897). U.S. Patent 4,448,885 and U.S. Patent 4,467,036 both disclose the expression of Br. crystal protein in E. coli. U.S. Patent 4,853,331 discloses Br. thuringiensis strain san diego (a.k.a. Br. tenebrionis, a.k.a. M-7) which can be used to control coleopteran pests in various environments. U.S. Patent No. 4,849,217 discloses Bacillus thuringiensis isolates active against the alfalfa weevil. One of the isolates disclosed is B. thuringiensis PS86A1 (NRRL B-18400).

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Brief Summary of the Invention

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The subject invention concerns a novel process for controlling lepidopteran pests. This process results from the unexpected discovery that certain coleopteran-active B.t. isolates also have activity against lepidopteran pests, e.g., the diamondback moth (Phutella xylostella). This discovery was particularly surprising because known coleopteran-active isolates such as Bacillus thuringiensis var. tenebrionis (Krieg et al., supra) (hereinafter referrred to as M-7) are not toxic to Lepidoptera.

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More specifically, the subject invention process uses B.t. microbes, or variants thereof, and/or their toxins, to control lepidopteran pests. Specific B.t. microbes useful according to the invention are B.t. PS86A1, B.t. PS50C, and B.t. PS43F. Further, the subject invention also includes the use of variants of the B.t. isolates of the invention which have substantially the same lepidopteran-active properties as the specifically exemplified B.t. isolates. Procedures for making mutants are well known in the microbiological art. Ultraviolet light and nitrosoguanidine are used extensively toward this end.

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The subject invention also includes the use of genes from the B.t. isolates of the invention which genes encode the lepidopteran-active toxins.

Still further, the invention also includes the treatment of substantially intact *B.t.* cells, and recombinant cells containing the genes of the invention, to prolong the lepidopteran activity when the substantially intact cells are applied to the environment of a target pest. Such treatment can be by chemical or physical means, or a combination of chemical and physical means, so long as the technique does not deleteriously affect the properties of the pesticide, nor diminish the cellular capability in protecting the pesticide. The treated cell acts as a protective coating for the pesticidal toxin. The toxin becomes available to act as such upon ingestion by a target insect.

Finally, the subject invention further concerns plants which have been transformed with genes encoding lepidopteran-active torins.

Brief Description of the Sequences

SEQ ID NO. 1 is the nucleotide sequence (open reading frame only) of the gene designated 50C.

SEQ ID NO. 2 is the predicted amino acid sequence of the toxin 50C.

SEQ ID NO. 3 is the composite nucleotide and amino acid sequences of the gene designated 43F.

SEQ ID NO. 4 is the predicted amino acid sequence of the toxin 43F.

SEQ ID NO. 5 is the nucleotide sequence (open reading frame only) of the gene designated 86A1.

SEQ ID NO. 6 is the predicted amino acid sequence of the toxin 86A1.

SEQ ID NO. 7 is an oligonucleotide probe designated 86A1-A.

25 <u>Detailed Disclosure of the Invention</u>

The Bacillus thuringiensis isolates useful according to the subject invention have the following characteristics in their biologically pure form:

Characteristics of Rt PS50C

Colony morphology--Large colony, dull surface, typical B.t.

Vegetative cell morphology-typical R.t.

Culture methods-typical for B.t.

Plagellar serotyping--PS50C belongs to serotype 18, kumamotoensis.

Crystal morphology-a sphere.

RFLP analysis-Southern hybridization of total DNA distinguishes B.t. PS50C from B.t.s.d. and other B.t. isolates.

Alkali-soluble proteins--SDS polyacrylamide gel electrophoresis (SDS-PAGE) shows a 130 kDa doublet protein.

The characteristics of B.t. PS86A1 with regard to colony morphology, vegetative cell morphology and culture methods are as given above for B.t. PS50C. However, these isolates differ, as shown in Table 1, with respect to inclusions, scrotype, and molecular weights of toxins.

B.t. PS43F is disclosed in U.S. Patent 4,996,155.

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A comparison of the characteristics of the B. thuringiensis strains of the subject invention to the characteristics of the known B.t. strains B. thuringiensis var. tenebrionis (M-7) and B. thuringiensis var. kurstaki (HD-1) is shown in Table 1.

	omparison of B.t. PS50 B.t. PS50C	B.t. PS86A1	B.t. PS43F	Rt HD-1	M-7
Inclusions:		Multiple attached	Flat, pointed, ellipse, plus small inclusions	Bipyramid	Flat square
Approxima molecular vof proteins SDS-PAGI	wt. doublet by	58,000 45,000	75,000 68,000 61,000	130,000 68,000	72,000 64,000
Serotype		sis wuhenensis	tolworthi	kurstaki	morrisoni

20 B.r. isolates useful according to the subject invention have been deposited. Also deposited are recombinant microbes comprising the B.r. genes of interest.

	Culture	Accession No	mber Deposit Date
	Bacillus thuringiensis PS50C	NRRL B-18746	January 9, 1991
25	E. coli NM522(pMYC1638) Bacillus thuringiensis PS86A1	NRRL B-18751	January 11, 1991
		NRRL B-18400	August 16, 1988
	E. coli NM522(pMYC2320)	NRRL B-18769	February 14, 1991
	Bacillus thuringiensis PS43F	NRRL B-18298	February 2, 1988
	E. coli XL1-Blue (pM1,98-4)	NRRL B-18291	January 15, 1988

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The cultures are on deposit in the permanent collection of the Northern Research Laboratory, U.S. Department of Agriculture, Peoria, IL, USA.

The subject cultures have been deposited under conditions that assure that access to the cultures will be available during the pendency of this patent application to one determined by the Commissioner of Patents and Trademarks to be entitled thereto under 37 CFR 1.14 and 35 USC 122. The deposits are available as required by foreign patent laws in countries wherein

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counterparts of the subject application, or its progeny, are filed. However, it should be understood that the availability f a deposit does not constitute a license to practice the subject invention in derogation of patent rights granted by governmental action.

Further, the subject culture deposits will be stored and made available to the public in accord with the provisions of the Budapest Treaty for the Deposit of Microorganisms, i.e., they will be stored with all the care necessary to keep them viable and uncontaminated for a period of at least five years after the most recent request for the furnishing of a sample of the deposit, and in any case, for a period of at least 30 (thirty) years after the date of deposit or for the enforceable life of any patent which may issue disclosing the cultures. The depositor acknowledges the duty to replace the deposits should the depository be unable to furnish a sample when requested, due to the condition of the deposit(s). All restrictions on the availability to the public of the subject culture deposits will be irrevocably removed upon the granting of a patent disclosing them.

The lepidopteran toxin genes of the subject invention can be isolated by known procedures and can be be introduced into a wide variety of microbial hosts. Expression of the toxin gene results, directly or indirectly, in the intracellular production and maintenance of the pesticide. With suitable hosts, e.g., *Pseudomonas*, the microbes can be applied to the situs of lepidopteran insects where they will proliferate and be ingested by the insects. The result is a control of the unwanted insects. Alternatively, the microbe hosting the toxin gene can be treated under conditions that prolong the activity of the toxin produced in the cell. The treated cell then can be applied to the environment of target pest(s). The resulting product retains the toxicity of the *B.t.* toxin.

Where the Rt toxin gene is introduced via a suitable vector into a microbial host, and said host is applied to the environment in a living state, it is important that certain host microbes be used. Microorganism hosts are selected which are known to occupy the "phytosphere" (phylloplane, phyllosphere, rhizosphere, and/or rhizoplane) of one or more crops of interest. These microorganisms are selected so as to be capable of successfully competing in the particular environment (crop and other insect habitats) with the wild-type microorganisms, provide for stable maintenance and expression of the gene expressing the polypeptide pesticide, and, desirably, provide for improved protection of the pesticide from environmental degradation and inactivation.

A large number of microorganisms are known to inhabit the phylloplane (the surface of the plant leaves) and/or the rhizosphere (the soil surrounding plant roots) of a wide variety of important crops. These microorganisms include bacteria, algae, and fungi. Of particular interest are microorganisms, such as bacteria, e.g., genera Pseudomonas, Erwinia, Serratia, Klebsiella, Xanthomonas, Streptomyces, Rhizobium, Rhodopseudomonas, Methylophilius, Agrobacterium, Acetobacter, Lactobacillus, Arthrobacter, Azotobacter, Leuconostoc, and Alcaligenes; fungi, particularly yeast, e.g., genera Saccharomyces, Cryptococcus, Kluyveromyces, Sporobolomyces,

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Rhodotorula, and Aureobasidium. Of particular interest are such phytosphere bacterial species as Pseudomonas syringae, Pseudomonas fluorescens, Serratia marcescens, Acetobacter xylinum, Agrobacterium tumefaciens, Rhodopseudomonas spheroides, Xanthomonas campestris, Rhizobium melioti, Alcaligenes entrophus, and Azotobacter vinlandii; and phytosphere yeast species such as Rhodotorula rubra, R. giutinis, R. marina, R. aurantiaca, Cryptococcus albidus, C. diffiuens, C. laurentil, Saccharomyces rosei, S. pretoriensis, S. cerevisiae, Sporobolomyces roseus, S. odorus, Khoveromyces veronae, and Aureobasidium pollulans. Of particular interest are the pigmented microorganisms.

A wide variety of ways are available for introducing the Br. gene expressing the toxin into the microorganism host under conditions which allow for stable maintenance and expression of the gene. One can provide for DNA constructs which include the transcriptional and translational regulatory signals for expression of the toxin gene, the toxin gene under their regulatory control and a DNA sequence homologous with a sequence in the host organism, whereby integration will occur, and/or a replication system which is functional in the host, whereby integration or stable maintenance will occur.

The transcriptional initiation signals will include a promoter and a transcriptional initiation start site. In some instances, it may be desirable to provide for regulative expression of the toxin, where expression of the toxin will only occur after release into the environment. This can be achieved with operators or a region binding to an activator or enhancers, which are capable of induction upon a change in the physical or chemical environment of the microorganisms. For example, a temperature sensitive regulatory region may be employed, where the organisms may be grown up in the laboratory without expression of a toxin, but upon release into the environment, expression begins. Other techniques may employ a specific nutrient medium in the laboratory, which inhibits the expression of the toxin, where the nutrient medium in the environment allows for expression of the toxin. For translational initiation, a ribosomal binding site and an initiation codon will be present.

Various manipulations may be employed for enhancing the expression of the messenger, particularly by using an active promoter, as well as by employing sequences, which enhance the stability of the messenger RNA. The initiation and translational termination region will involve stop codon(s), a terminator region, and optionally, a polyadenylation signal.

In the direction of transcription, namely in the 5' to 3' direction of the coding or sense sequence, the construct can involve the transcriptional regulatory region, if any, and the promoter, where the regulatory region may be either 5' or 3' of the promoter, the ribosomal binding site, the initiation codon, the structural gene having an open reading frame in phase with the initiation codon, the stop codon(s), the polyadenylation signal sequence, if any, and the terminator region. This sequence as a double strand may be used by itself for transformation of a microorganism host, but will usually be included with a DNA sequence involving a marker, where the second

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DNA sequence may be joined to the toxin expression construct during introduction of the DNA into the host.

which have been modified or transformed. The marker will normally provide for selective advantage, for example, providing for biocide resistance, e.g., resistance to antibiotics or heavy metals; complementation, so as to provide prototropy to an auxotrophic host, or the like. Preferably, complementation is employed, so that the modified host may not only be selected, but may also be competitive in the field. One or more markers may be employed in the development of the constructs, as well as for modifying the host. The organisms may be further modified by providing for a competitive advantage against other wild-type microorganisms in the field. For example, genes expressing metal chelating agents, e.g., siderophores, may be introduced into the host along with the structural gene expressing the toxin. In this manner, the enhanced expression of a siderophore may provide for a competitive advantage for the toxin-producing host, so that it may effectively compete with the wild-type microorganisms and stably occupy a niche in the environment.

Where stable episomal maintenance or integration is desired, a plasmid will be employed which has a replication system which is functional in the host. The replication system may be derived from the chromosome, an episomal element normally present in the host or a different host, or a replication system from a virus which is stable in the host. A large number of plasmids are available, such as pBR322, pACYC184, RSF1010, pRO1614, and the like. See for example, Olson et al. (1982) J. Bacteriol. 150:6069, and Bagdasarian et al. (1981) Gene 16:237, and U.S. Patent Nos. 4,356,270, 4,362,817, and 4,371,625.

Where no functional replication system is present, the construct will also include a sequence of at least 50 basepairs (bp), preferably at least about 100 bp, and usually not more than about 1000 bp of a sequence homologous with a sequence in the host. In this way, the probability of legitimate recombination is enhanced, so that the gene will be integrated into the host and stably maintained by the host. Desirably, the toxin gene will be in close proximity to the gene providing for complementation as well as the gene providing for the competitive advantage. Therefore, in the event that a toxin gene is lost, the resulting organism will be likely to also lose the complementing gene and/or the gene providing for the competitive advantage, so that it will be unable to compete in the environment with the gene retaining the intact construct.

A large number of transcriptional regulatory regions are available from a wide variety of microorganism hosts, such as bacteria, bacteriophage, cyanobacteria, algae, fungi, and the like. Various transcriptional regulatory regions include the regions associated with the *trp* gene, *lac* gene, *gal* gene, the lambda left and right promoters, the *tac* promoter, and the naturally-occurring promoters associated with the toxin gene, where functional in the host. See for example, U.S. Patent Nos. 4,332,898, 4,342,832 and 4,356,270. The termination region may be the termination

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region normally associated with the transcriptional initiation region or a different transcriptional initiation region, so long as the two regions are compatible and functional in the host.

The B.t. gene can be introduced between the transcriptional and translational initiation region and the transcriptional and translational termination region, so as to be under the regulatory control of the initiation region. This construct can be included in a plasmid, which could include at least one replication system, but may include more than one, where one replication system is employed for cloning during the development of the plasmid and the second replication system is necessary for functioning in the ultimate host. In addition, one or more markers may be present, which have been described previously. Where integration is desired, the plasmid will desirably include a sequence homologous with the host genome.

The transformants can be isolated in accordance with conventional ways, usually employing a selection technique, which allows for selection of the desired organism as against unmodified organisms or transferring organisms, when present. The transformants then can be tested for pesticidal activity.

Suitable host cells, where the pesticide-containing cells will be treated to prolong the activity of the toxin in the cell when the then treated cell is applied to the environment of target pest(s), may include either prokaryotes or eukaryotes, normally being limited to those cells which do not produce substances toxic to higher organisms, such as mammals. However, organisms which produce substances toxic to higher organisms could be used, where the toxin is unstable or the level of application sufficiently low as to avoid any possibility of toxicity to a mammalian host. As hosts, of particular interest will be the prokaryotes and the lower eukaryotes, such as fungi. Illustrative prokaryotes, both Gram-negative and -positive, include Enterobacteriaceae, such as Escherichia, Erwinia, Shigella, Salmonella, and Proteus, Bacillaceae; Rhizobiceae, such as Rhizobium; Spirillaceae, such as photobacterium, Zymomonas, Serratia, Aeromonas, Vibrio, Desulfovibrio, Spirillum; Lactobacillaceae; Pseudomonadaceae, such as Pseudomonas and Acetobacter, Azotobacteraceae and Nitrobacteraceae. Among eukaryotes are fungi, such as Phycomycetes and Ascomycetes, which includes yeast, such as Saccharomyces and such as Rhodotorula, Aureobasidium, Schizosaccharomyces; and Basidiomycetes yeast, Sporobolomyces, and the like.

The cell will usually be intact and be substantially in the proliferative form when treated, rather than in a spore form, although in some instances spores may be employed.

Treatment of the microbial cell, e.g., a microbe containing the B.r. lepidopteran toxin gene, can be by chemical or physical means, or by a combination of chemical and/or physical means, so long as the technique does not deleteriously affect the properties of the toxin, nor diminish the cellular capability in protecting the toxin. Examples of chemical reagents are halogenating agents, particularly halogens of atomic no. 17-80. More particularly, iodine can be used under mild conditions and for sufficient time to achieve the desired results. Other suitable

techniques include treatment with aldehydes, such as formaldehyde and glutaraldehyde; antiinfectives, such as zephiran chloride and cetylpyridinium chloride; alcohols, such as isopropyl and
ethanol; various histologic firatives, such as Bouin's fixative and Helly's fixative (See: Humason,
Gretchen L. [1967] Animal Tissue Techniques, W.H. Freeman and Company); or a combination
of physical (heat) and chemical agents that preserve and prolong the activity of the toxin produced
in the cell when the cell is administered to the host animal. Examples of physical means are short
wavelength radiation such as gamma-radiation and X-radiation, freezing, UV irradiation,
lyophilization, and the like.

The cells generally will have enhanced structural stability which will enhance resistance to environmental conditions. Where the pesticide is in a proform, the method of inactivation should be selected so as not to inhibit processing of the proform to the mature form of the pesticide by the target pest pathogen. For example, formaldehyde will crosslink proteins and could inhibit processing of the proform of a polypeptide pesticide. The method of inactivation or killing retains at least a substantial portion of the bio-availability or bioactivity of the toxin.

Characteristics of particular interest in selecting a host cell for purposes of production include ease of introducing the B.t. gene into the host, availability of expression systems, efficiency of expression, stability of the pesticide in the host, and the presence of auxiliary genetic capabilities. Characteristics of interest for use as a pesticide microcapsule include protective qualities for the pesticide, such as thick cell walls, pigmentation, and intracellular packaging or formation of inclusion bodies; leaf affinity; lack of mammalian toxicity; attractiveness to pests for ingestion; ease of killing and fixing without damage to the toxin; and the like. Other considerations include ease of formulation and handling, economics, storage stability, and the like.

Host organisms of particular interest include yeast, such as Rhodotorula sp., Aureobasidium sp., Saccharomyces sp., and Sporobolomyces sp.; phylloplane organisms such as Pseudomonas sp., Erwinia sp. and Flavobacterium sp.; or such other organisms as Escherichia, Lactobacillus sp., Bacillus sp., and the like. Specific organisms include Pseudomonas aeruginosa, Pseudomonas fluorescens, Saccharomyces cerevisiae, Bacillus thuringiensis, Escherichia coli, Bacillus subtilis, and the like.

The cellular host containing the Br. lepidopteran gene may be grown in any convenient nutrient medium, where the DNA construct provides a selective advantage, providing for a selective medium so that substantially all or all of the cells retain the Br. gene. These cells may then be harvested in accordance with conventional ways. Alternatively, the cells can be treated prior to harvesting.

The B.t. cells may be formulated in a variety of ways. They may be employed as wettable powders, granules or dusts, by mixing with various inert materials, such as inorganic minerals (phyllosilicates, carbonates, sulfates, phosphates, and the like) or botanical materials (powdered corncobs, rice hulls, wainut shells, and the like). The formulations may include spreader-sticker

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adjuvants, stabilizing agents, other pesticidal additives, or surfactants. Liquid formulations may be aqueous-based or non-aqueous and employed as foams, gels, suspensions, emulsifiable concentrates, or the like. The ingredients may include theological agents, surfactants, emulsifiers, dispersants, or polymers.

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The pesticidal concentration will vary widely depending upon the nature of the particular formulation, particularly whether it is a concentrate or to be used directly. The pesticide will be present in at least about 1% by weight and may be about 100% by weight. The dry formulations will have from about 1-95% by weight of the pesticide while the liquid formulations will generally be from about 1-60% by weight of the solids in the liquid phase. The formulations will generally have from about 10² to about 10⁴ cells/mg. These formulations will be administered at about 50 mg (liquid or dry) to 1 kg or more per hectare.

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The formulations can be applied to the environment of the lepidopteran pest(s), e.g., plants, soil or water, by spraying, dusting, sprinkling, or the like.

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Following are examples which illustrate procedures, including the best mode, for practicing the invention. These examples should not be construed as limiting. All percentages are by weight and all solvent mixture proportions are by volume unless otherwise noted.

Example 1 - Culturing Bacillus thuringiensis Isolates

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A subculture of a B.t. isolate of the invention can be used to inoculate the following medium, a peptone, glucose, salts medium.

Bacro Pentone	7.50 g/I
	1.00 g/I
•	3.40 g/l
-	4.35 g/l
_	5.00 ml/l
CaCl ₂ Solution	5.00 ml/t
	Bacto Peptone Glucose KH ₂ PO ₄ K ₂ HPO ₄ Salt Solution

pH 7.2

	Salts Solution (100 ml)	
	MgSO ₄ 7H ₂ O	2.46 g
	MnSO ₄ H ₂ O	0.04 g
	ZnSO ₄ 7H ₂ O	0.28 g
5	FeSO ₄ ·7H ₂ O	0.40 g

CaCl₂ Solution (100 ml)

CaCl₂·2H₂O

3.66 g

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The salts solution and CaCl₂ solution are filter-sterilized and added to the autoclaved and cooked broth at the time of inoculation. Flasks are incubated at 30°C on a rotary shaker at 200 rpm for 64 hr.

The above procedure can be readily scaled up to large fermentors by procedures well known in the art.

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The R.z. spores and crystals, obtained in the above fermentation, can be isolated by procedures well known in the art. A frequently-used procedure is to subject the harvested fermentation broth to separation techniques, e.g., centrifugation.

Example 2 - Cloning of a Toxin Gene from B.z. Isolate PS50C

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Total cellular DNA was prepared from *Bacillus thuringlensis* (*B.t.*) cells grown to an optical density, at 600 nm, of 1.0. The cells were recovered by centrifugation and protoplasts were prepared in TES buffer (30 mM Tris-HCl, 10 mM EDTA, 50 mM NaCl, pH = 8.0) containing 20% sucrose and 50 mg/ml lysozyme. The protoplasts were lysed by addition of SDS to a final concentration of 4%. The cellular material was precipitated overnight at 4°C in 100 mM (final concentration) neutral potassium chloride. The supernate was extracted twice with phenol/chloroform (1:1). Nucleic acids were precipitated with ethanol and DNA was purified by isopycnic banding on cesium chloride-ethidium bromide gradients.

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Total cellular DNA from B.r. subsp. kumamotoensis (B.r. kum.), isolate PSSOC, was digested with HindIII and fractionated by electrophoresis on a 0.8% (w/v) agarose-TAE (50 mM Tris-HCl, 20 mM NaOAc, 2.5 mM EDTA, pH = 8.0) buffered gel. A Southern blot of the gel was hybridized with a [32P]-radiolabeled oligonucleotide probe. Results showed that the hybridizing fragments of PSSOC are approximately 12 kb and 1.7 kb in size.

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A library was constructed from PS50C total cellular DNA partially digested with Sau3A and size fractionated by gel electrophoresis. The 9-23 kb region of the gel was excised and the DNA was electrocluted and then concentrated using an Elutip-dTM ion exchange column (Schleicher and Schuel, Keene, NH). The isolated Sau3A fragments were ligated into BamHI-digested LambdaGEM-11TM (PROMEGA). The packaged phage were plated on E. coli KW251

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cells (PROMEGA) at a high titer and screened using the radiolabeled oligonucleotide probe. Hybridizing plaques were purified and rescreened at a lower plaque density. Single isolated, purified plaques that hybridized with the probe were used to infect E. coli KW251 cells in liquid culture for preparation of phage for DNA isolation. DNA was isolated by standard procedures. Preparative amounts of DNA were digested with XhoI (to release the inserted DNA from lambda sequences) and separated by electrophoresis on a 0.6% agarose-TAE gel. The large fragments were purified by ion exchange chromatography as above and ligated to Xhol-digested, dephosphorylated pHTBlueII (an E. coli/B. thuringiensis shuttle vector comprised of pBluescript s/k [Stratagene] and the replication origin from a resident B.t. plasmid [D. Lereclus et al. [1989] FEMS Microbiology Letters 60:211-218]). The ligation mix was introduced by transformation into competent E. coli NM522 cells (ATCC 47000) and plated on LB agar containing ampicillin, isopropyl- (β) -D-thiogalactoside (IPTG) and 5-bromo-4-chloro-4-indolyl- (β) -D-galactoside (XGAL). White colonies, with putative restriction fragment insertions in the (β) -galactosidase gene of pHTBlueII, were subjected to standard rapid plasmid purification procedures. Plasmids were analyzed by Xhol digestion and agarose gel electrophoresis. The desired plasmid construct, pMYC1638, contains an approximately 12 kb Xhol insert. The nucleotide sequence (open reading frame only) is shown in SEQ ID NO. 1. The predicted amino acid sequence of the toxin is shown in SEQ ID NO. 2.

Plasmid pMYC1638 was introduced into an acrystalliferous (Cry) R.t. host (HD-1 cryB obtained from A. Aronson, Purdue University) by electroporation. Expression of an approximately 130 kDa protein was verified by SDS-PAGE.

Plasmid pMYC1638 containing the B.t. toxin gene, can be removed from the transformed host microbe by use of standard well-known procedures. For example, E. coli NM522[pMYC1638] NRRL B-18751 can be subjected to cleared lysate isopycnic density gradient procedures, and the like, to recover pMYC1638.

Example 3 - Cloning of Toxin Gene From Rt. Isolate PS43F and Transformation into Pseudomonas

Total cellular DNA was prepared by growing the cells of B.t. isolate PS43F and M-7 to a low optical density ($OD_{600} = 1.0$) and recovering the cells by centrifugation. The cells were protoplasted in a buffer containing 20% sucrose and 50 mg/ml lysozyme. The protoplasts were lysed by addition of SDS to a final concentration of 4%. The cellular material was precipitated overnight at 4°C in 100 mM neutral potassium chloride. The supernate was phenol/chloroform extracted twice and the DNA precipitated in 68% ethanol. The DNA was purified on a cesium chloride gradient. DNAs from strains 43F and M-7 (as a standard of reference) were digested with EcoRI and run out on a 0.8% agarose gel. The gel was Southern blotted and probed with the nick translated ORF KmnI to PstI fragment of the toxin encoding gene isolated from M-7 (this

will be subsequently referred to as Probe). The results showed 43F to hybridize to Probe at 7.5 kb which is different than the standard.

Preparative amounts of 43F DNA were digested with EcoRI and run out on a 0.8% agarose gel. The 7.5 kb region of the preparative gel was isolated and the DNA electrocluted and concentrated using an ELUTIPTM-d (Schleicher and Schuell, Keene, NH) ion exchange column. A sample was blotted and probed to verify the fragment was indeed isolated. The 7.5 kb EcoRI fragment was ligated to Lambda ZAPTM EcoRI arms. The packaged recombinant phage were plated out with E. coli strain BB4 (Stratagene Cloning Systems, La Jolla, CA) to give high plaque density.

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The plaques were screened by standard procedures with Probe. The plaques that hybridized were purified and re-screened at a lower plaque density. The resulting phage were grown with M13 helper phage (Stratagene) and the recombinant BLUESCRIPTTM plasmid was automatically excised and packaged. The "phagemid" was re-infected in XL1-blue E coli cells (Stratagene) as part of the automatic excision process. The infected XL1-blue cells were screened for ampicillin resistance and the resulting colonies were miniprepped to find the desired plasmid pM1,98-4. The recombinant E coli XL1-Blue (pM1,98-4) strain is called MR381.

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The plasmid pM1,98-4 contained a 7.5 kb EcoRI insert. To verify that this insert was the one of interest, a Southern blot was performed and probed. The 7.5 kb band hybridized with Probe, confirming that the fragment had been cloned. Restriction endonuclease analysis of the 7.5 kb EcoRI fragment with the enzymes HindIII, Pstl, Spel, BamHI and XbaI was done to show that a gene different from M-7 had been cloned. The enzymes which cut inside the 7.5 kb EcoRI fragment were HindIII (twice) SpeI (twice) and PstI (once). The open reading frame (ORF) of the 43F gene cut once with HindIII, twice with SpeI and did not cut with XbaI, EcoRI, or BamHI. Sequence data showed an open reading frame of 1963 bp with at best 70% sequence similarity to the toxin encoding gene of M-7.

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The cloned toxin gene from PS43F can be modified for expression in *P. fluorescens* in the following way:

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(1) A plasmid containing the Ptac-promoted cryIA(b)-like toxin gene can be made using a 3-way ligation involving the Ptac promoter and toxin gene on a BamHI-PstI fragment of about 4500 bp from pM3,130-7 (from MR420, NRRL B-18332, disclosed in U.S. Patent No. 5,055,294), a NotI-BamHI fragment of about 5500 bp from pTJS260 (containing the tetracycline resistance genes, available from Dr. Donald Helinski, U.C. San Diego), and a NotI-PstI fragment of about 6100 bp from pTJS260 (containing the replication region). The assembled plasmid is recovered following transformation of E. coli and growth under tetracycline selection.

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(2) A plasmid containing the Ptac-promoted 43F toxin gene can be made by ligating the toxin gene-containing Fspl-Sspl fragment of about 2200 bp from pM1,98-4 (from MR381(pM1,98-4), NRRL B-18291) into the Smal site of the E. coli vector, pKK223-3 (Pharmacia). The Ptac-

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promoted 43F toxin plasmid can be recovered following transformation of *E. coli*, growth under ampleillin selection, and screening for plasmids with inserts in the proper rientation for expression from the *tac* promoter by techniques well known in the art.

(3) The Ptac-promoted 43F toxin can be assembled into, for example, the pTJS260-derived vector in a three-way ligation using the 12.6 kb DNA fragment having BamHI and filled-in NsiI ends from the plasmid resulting from step 1 above, to the BamHI-NsiI Ptac-containing fragment of about 1.2 kb and the NsiI-ScaI fragment of about 2.1 kb containing the 3' end of the 43F toxin gene and adjacent vector DNA from the plasmid resulting from step 2 above.

The resulting pTJS260-derived 43F toxin expression plasmid can be introduced into Pseudomonas fluorescens by electroporation.

The above cloning procedures were conducted using standard procedures unless otherwise noted.

The various methods employed in the preparation of the plasmids and transformation of host organisms are well known in the art. These procedures are described in Maniatis, T., E.F. Fritsch, J. Sambrook (1982) Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, New York. Thus, it is within the skill of those in the genetic engineering art to extract DNA from microbial cells, perform restriction enzyme digestions, electrophorese DNA fragments, tail and anneal plasmid and insert DNA, ligate DNA, transform cells, prepare plasmid DNA, electrophorese proteins, and sequence DNA.

The restriction enzymes disclosed herein can be purchased from Boehringer Mannheim, Indianapolis, IN, or New England BioLabs, Beverly, MA. The enzymes were used according to the instructions provided by the supplier.

Plasmid pM1,98-4 containing the B.t. toxin gene, can be removed from the transformed host microbe by use of standard well-known procedures. For example, E. coli XL1-Blue (pM1,98-4) can be subjected to cleared lysate isopycnic density gradient procedures, and the like, to recover pM1,98-4.

Example 4 - Molecular Cloning of Gene Encoding a Toxin from Bacillus thuringiensis Strain PS86A1

Total cellular DNA was prepared from PS86A1 cells grown to an optical density, at 600 nm, of 1.0. Cells were pelleted by centrifugation and resuspended in protoplast buffer (20 mg/ml lysozyme in 0.3 M sucrose, 25 mM Tris-Cl, pH 8.0, 25 mM EDTA). After incubation at 37°C for 1 hour, protoplasts were lysed by two cycles of freezing and thawing. Nine volumes of a solution of 0.1 M NaCl, 0.1% SDS, 0.1 M Tris-Cl were added to complete lysis. The cleared lysate was extracted twice with phenol:chloroform (1:1). Nucleic acids were precipitated with two volumes of ethanol and pelleted by centrifugation. The pellet was resuspended in 10 mM Tris-Cl, 1 mM EDTA (TE), pH 8.0, and RNAse was added to a final concentration of 50 μ g/ml. After

incubation at 37°C for 1 hour, the solution was extracted once each with phenol:chloroform (1:1) and TE-saturated chloroform. DNA was precipitated from the aqueous phase by the addition of one-tenth volume of 3 M NaOAc and two volumes of ethanol. DNA was pelleted by centrifugation, washed with 70% ethanol, dried, and resuspended in TE.

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Restriction fragment length polymorphism (RFLP) analyses were performed by standard hybridization of southern blots of PS86A1 DNA with a ³²P-labeled oligonucleotide probe designated as 86A1-A. The sequence of the 86A1-A probe was:

5' ATG ATT GAT TCT AAA ACA ACA TTA CCA AGA CAT TCT/A TTA ATT/A CAT ACT/A ATT/A AA 3' (SEQ ID NO. 7)

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The probe was mixed at four positions, as shown. Hybridizing bands included an approximately 3.6 kbp *HindIII* fragment and an approximately 9.3 kbp *EcoRV* fragment.

restriction digests were fractionated by agarose gel electrophoresis. DNA fragments 6.6 to 23 kbp

A gene library was constructed from PS86A1 DNA partially digested with Sau3A. Partial

in size were excised from the gel, electroeluted from the gel slice, and recovered by ethanol precipitation after purification on an Elutip-D ion exchange column. The Sau3A inserts were ligated into BamHI-digested LambdaGem-11 (Promega, Madison, WI). Recombinant phage were packaged and plated on E. coli KW251 cells (Promega). Plaques were screened by hybridization with the radiolabeled 86A1-A oligonucleotide probe. Hybridizing phage were plaque-purified and

used to infect liquid cultures of *E. coli* KW251 cells for isolation of phage DNA by standard procedures (Maniatis et al. [1982] *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY). For subcloning, preparative amounts of DNA were digested with *EcoRI* and *SalI*, and electrophoresed on an agarose gel. The approximately 2.9 kbp

band containing the toxin gene was excised from the gel, electroeluted from the gel slice, and purified by ion exchange chromatography as above. The purified DNA insert was ligated into EcoRI + Sall-digested pHTBlueII (an E. coli/B.t. shuttle vector comprised of pBlueScript S/K, Stratagene, San Diego, CA) and the replication origin from a resident B.t. plasmid (D. Lereclus et al. [1989] FEMS Microbiol. Lett. 60:211-218). The ligation mix was used to transform frozen,

competent E. coli NM522 cells (ATCC 47000). Transformants were plated on LB agar (Maniatis et al., supra) containing ampicillin, isopropyl-(β)-D-thiogalactoside (IPTG), and 5-bromo-4-chloro-3-indolyl-(β)-D-galactoside (XGAL). Plasmids were purified from putative recombinants by

alkaline lysis (Maniatis et al., supra) and analyzed by electrophoresis of EcoRI and Sall digests on agarose gels. The desired plasmid construct, pMYC2320, contains the toxin gene of the invention.

The DNA sequence of this gene is shown in SEQ ID NO. 5. The toxin expressed by this gene is shown in SEQ ID NO. 6.

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Plasmid pMYC2320 was introduced into an acrystalliferous (Cry \(^\) &t. host (&t. HD-1). Cry B, A.I. Aronson, Purdue University, West Lafayatte, IN) by electroporation. Expression of an approximately 58 kDa protein is verified by SDS-PAGE analysis.



Plasmid pMYC2320 containing the B.t. toxin gene, can be removed from the transformed host microbe by use of standard well-known procedures. For example, E coli NM522(pMYC2320) can be subjected to cleared lysate isopycnic density gradient procedures, and the like, to recover pMYC2320.

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Example 5 - 43F Toxin Production by a Transformed Pseudomonas fluorescens Host

A transformed Pseudomonas fluorescens containing the PS43F gene was grown in the following medium using a 1% inoculum grown in LB medium with 30 μ g/ml tetracycline:

aoaic.	Glycerol	65 g/L
10	Na citrate 2H ₂ O	7.14
10	нст	20
	Amberex 1003	20
	NaNO ₃	5
	(NH ₄) ₂ SO ₄	2.3
15	32°C at 300 rpm.	

These were 72 hour fermentations with induction and supplementation taking place at 24 hours. They were induced with 2 mM IPTG and supplemented with the following:

	24 1100101 2311	Amisoy	20.0 g/L
-		MgSO ₄ -7H ₂ O	0.4
20		K ₂ HPO ₄	1.6
		KCI	1.6

The toxin concentration can be determined using laser densitometry (LKB) to quantify the approximately 70 kDa toxin protein found in the *P. fluorescens* host after Coomassie staining of polyacrylamide gels containing SDS.

Example 6 - Testing of B.t. Toxins Against the Diamondback Moth

- (A) A spore crystal preparation of a B.t. clone comprising the PS86A1 gene was toxic to the lepidopteran pest, diamondback moth *Plutella xylostella*, in a 1.5% agar artificial diet assay. The B.t. clone was grown as disclosed in Example 1. Rates greater than 100 microgram protein/gram diet gave 100% control of this pest in 6 days.
- (B) A spore crystal preparation of a Rt. clone comprising the PS50C gene was toxic to the lepidopteran pest, diamondback moth, in a 1.5% agar artificial diet assay. The Rt. clone was grown as disclosed in Example 1. Rates greater than 100 microgram protein/gram diet gave 100% control of this pest in 6 days.

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(C) A Pseudomonas fluorescens clone comprising the PS43F gene was toxic to the lepidopteran pest, diamondback moth, in a 1.5% agar artificial diet assay. Rates greater than 40 microgram protein/gram diet gave 100% control of this pest in 6 days.

Example 7 - Further Testing of Rt. Toxins Against the Diamondback Moth

Toxins of the subject invention were produced by recombinant cells which had been transformed with genes according to the subject invention. The toxins produced by the recombinant cells were then tested for their activity against diamondback moths. The results of these experiments are shown in Table 2. These experiments were conducted as described in Example 6.

	Table 2						
Source strain	Cloned Toxin Gene	Host	Clone	Diamondback Moth LC ₅₀ (µg toxin/g diet)			
PS86A1	86A1	B. thuringiensis	MR506	79			
PS50C	50C	B. thuringiensis	MR505	19			
PS43F	43F	P. fluorescens	MR816	. 11			

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Example 8 - Insertion of Toxin Genes Into Plants

One aspect of the subject invention is the transformation of plants with genes encoding a lepidopteran toxin. The transformed plants are resistant to attack by lepidopterans.

Genes encoding lepidopteran-active toxins, as disclosed herein, can be inserted into plant cells using a variety of techniques which are well known in the art. For example, a large number of cloning vectors comprising a replication system in *E. coli* and a marker that permits selection of the transformed cells are available for preparation for the insertion of foreign genes into higher plants. The vectors comprise, for example, pBR322, pUC series, M13mp series, pACYC184, etc. Accordingly, the sequence encoding the *B.t.* toxin can be inserted into the vector at a suitable restriction site. The resulting plasmid is used for transformation into *E. coli*. The *E. coli* cells are cultivated in a suitable nutrient medium, then harvested and lysed. The plasmid is recovered. Sequence analysis, restriction analysis, electrophoresis, and other biochemical-molecular biological methods are generally carried out as methods of analysis. After each manipulation, the DNA sequence used can be cleaved and joined to the next DNA sequence. Each plasmid sequence can be cloned in the same or other plasmids. Depending on the method of inserting desired genes into the plant, other DNA sequences may be necessary. If, for example, the Ti or Ri plasmid is

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used for the transformation of the plant cell, then at least the right border, but often the right and the left border of the Ti or Ri plasmid T-DNA, has to be joined as the flanking region of the genes to be inserted.

The use of T-DNA for the transformation of plant cells has been intensively researched and sufficiently described in EP 120 516; Hoekema (1985) In: The Binary Plant Vector System, Offset-durkkerij Kapters B.V., Alblasserdam, Chapter 5; Fraley et al., Crit. Rev. Plant Sci. 4:1-46; and An et al. (1985) EMBO J. 4:277-287.

Once the inserted DNA has been integrated in the genome, it is relatively stable there and, as a rule, does not come out again. It normally contains a selection marker that confers on the transformed plant cells resistance to a biocide or an antibiotic, such as kanamycin, G 418, bleomycin, hygromycin, or chloramphenicol, inter alia. The individually employed marker should accordingly permit the selection of transformed cells rather than cells that do not contain the inserted DNA.

A large number of techniques are available for inserting DNA into a plant host cell. Those techniques include transformation with T-DNA using Agrobacterium tumefaciens or Agrobacterium rhizogenes as transformation agent, fusion, injection, or electroporation as well as other possible methods. If agrobacteria are used for the transformation, the DNA to be inserted has to be cloned into special plasmids, namely either into an intermediate vector or into a binary vector. The intermediate vectors can be integrated into the TI or Ri plasmid by homologous recombination owing to sequences that are homologous to sequences in the T-DNA. The Ti or Ri plasmid also comprises the vir region necessary for the transfer of the T-DNA. Intermediate vectors cannot replicate themselves in agrobacteria. The intermediate vector can be transferred into Agrobacterium tumefaciens by means of a helper plasmid (conjugation). Binary vectors can replicate themselves both in E. coli and in agrobacteria. They comprise a selection marker gene and a linker or polylinker which are framed by the right and left T-DNA border regions. They can be transformed directly into agrobacteria (Holsters et al. [1978] Mol. Gen. Genet. 163:181-187). The agrobacterium used as host cell is to comprise a plasmid carrying a vir region. The vir region is necessary for the transfer of the T-DNA into the plant cell. Additional T-DNA may be contained. The bacterium so transformed is used for the transformation of plant cells. Plant explants can advantageously be cultivated with Agrobacterium numefaciens or Agrobacterium rhizogenes for the transfer of the DNA into the plant cell. Whole plants can then be regenerated from the infected plant material (for example, pieces of leaf, segments of stalk, roots, but also protoplasts or suspension-cultivated cells) in a suitable medium, which may contain antibiotics or biocides for selection. The plants so obtained can then be tested for the presence of the inserted DNA. No special demands are made of the plasmids in the case of injection and electroporation. It is possible to use ordinary plasmids, such as, for example, pUC derivatives.

The transformed cells grow inside the plants in the usual manner. They can form germ cells and transmit the transformed trait(s) to progeny plants. Such plants can be grown in the normal manner and crossed with plants that have the same transformed hereditary factors or other hereditary factors. The resulting hybrid individuals have the corresponding phenotypic properties.

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Example 9 - Cloning of Novel Rt. Genes Into Insect Viruses

A number of viruses are known to infect insects. These viruses include, for example, baculoviruses and entomopoxviruses. In one embodiment of the subject invention, lepidopteranactive genes, as described herein, can be placed with the genome of the insect virus, thus enhancing the pathogenicity of the virus. Methods for constructing insect viruses which comprise B.t. toxin genes are well known and readily practiced by those skilled in the art. These procedures are described, for example, in Merryweather et al. (Merryweather, A.T., U. Weyer, M.P.G. Harris, M. Hirst, T. Booth, R.D. Possee [1990] J. Gen. Virol. 71:1535-1544) and Martens et al. (Martens, J.W.M., G. Honee, D. Zuidema, J.W.M. van Lent, B. Visser, J.M. Vlak [1990] Appl. Environmental Microbiol. 56(9):2764-2770).

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It should be understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and the scope of the appended claims.

SEQUENCE LISTING

THE PART OF THE PA	
(1) GENERAL INFORMATION:	
(i) APPLICANT: Uyeda, Kendrick A. Bradfisch, Gregory A.	
(ii) TITLE OF INVENTION: Process for Controlling Lepidopteran Pests	
(iii) NUMBER OF SEQUENCES: 7	
(iv) CORRESPONDENCE ADDRESS: (A) ADDRESSEE: David R. Saliwanchik (B) STREET: 2421 N.W. 41st Street, Suite A-1 (C) CITY: Gainesville (D) STATE: FL (E) COUNTRY: USA (F) ZIP: 32606	
(V) COMPUTER READABLE FORM: (A) MEDIUM TYPE: Floppy disk (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS (D) SOFTWARE: Patentin Release \$1.0, Version \$1.25	
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(viii) ATTORNEY/AGENT INFORMATION: (A) NAME: Saliwanchik, David R. (B) REGISTRATION NUMBER: 31,794	
(ix) TELECOMMUNICATION INFORMATION: (A) TELEPHONE: 904-375-8100 (B) TELEFAX: 904-372-5800	
(2) INFORMATION FOR SEQ ID NO:1:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 3471 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
(11) MOLECULE TYPE: DNA (genomic)	
(iii) HYPOTHETICAL: NO	
(iv) anti-sense; no	
(Vi) ORIGINAL SOURCE: (A) ORGANISM: Bacillus thuringiensis (B) STRAIN: kumamotoensis (C) INDIVIDUAL ISOLATE: PS50C	
(Vii) IMMEDIATE SOURCE: (B) CLONE: E. coli NM522(pMYC1638), NRRL B-18751	
(x1) SEQUENCE DESCRIPTION: SEQ ID NO:1:	
ATGRETCCAR ATRATCARRA TGRATATGRA ATTATAGRIG CGRCACCTIC TACATCIGIA 60	_
TCCAGFGATT CTARCAGATA CCCTTTTGCG ARTGAGCCAA CAGATGCGTT ACAAAATATG 120	,
ARTTATAAAG ATTATCTGAA AATGTCTGGG GGAGAGAATC CTGAATTATT TGGAAATCCG 180	i

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GAGACGTTTA	TTAGTTCATC	CACGATTCAA	ACTGGAATTG	GCATTGTTGG	TCGAATACTA	. 240
GGAGCTTTAG	GGGTTCCATT	TGCTAGTCAG	ATAGCTAGTT	TCTATAGTTT	CATTGTTGGT	. 300
CAATTATGGC	CGTCAAAGAG	CGTAGATATA	TGGGGAGAAA	TTATGGAACG	AGTGGAAGAA	360
CTCGTTGATC	aaaaaataga	AAAATATGTA	aaagataagg	CTCTTGCTGA	Attaaaaggg	420
CTAGGAAATG	CTTTGGATGT	ATATCAGCAG	TCACTTGAAG	ATTGGCTGGA	AAATCGCAAT	480
GATGCAAGAA	CTAGAAGTGT	TGTTTCTAAT	CAATTTATAG	CTTTAGATCT	TAACTTTGTT	540
AGTTCAATTC	CATCTTTTGC	AGTATCCGGA	CACGAAGTAC	TATTATTAGC	AGTATATGCA	600
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GGATTTACAC	CAGGTGAAAT	TTCTAGATTT	TATAATCGTC	AAGTGCAACT	TACCGCTGAA	720
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CGTCCGCCAC	ACTTGTTTGA	TATACTCAGC	TCAGTAGAAA	TTAATACAAG	TAGAGGGGGT	1080
ATTACGTTAA	ATAATGATGC	ATATATAAAC	TACTGGTCAG	GACATACCCT	AAAATATCGT	1140
AGAACAGCTG	ATTCGACCGT	AACATACACA	GCTAATTACG	GTCGAATCAC	TTCAGAAAAG	1200
AATTCATTTG	CACTTGAGGA	TAGGGATATT	TTTGAAATTA	ATTCAACTGT	GGCAAACCTA	1260
GCTAATTACT	ACCALAGGC	ATATGGTGTG	CCGGGATCTT	GGTTCCATAT	GGTAAAAAGG	1320
GGAACCTCAT	CAACAACAGC	GTATTTATAT	TCARABACAC	ATACAGCTCT	CCAAGGGTGT	1380
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AGCTATAGTC	ATAGATTATC	TCATATTACC	TCCCATTCTT	TCTCTAAAAA	TGGGAGTGCA	1500
TACTATGGGA	GTTTCCCTGT	atttgtttgg	ACACATACTA	GTGCGGATTT	AAATAATACA	1560
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	ATTTTAGCTC					1980
	CATATGAGGC					2040
					AGTAAATCAA	2100
					ACGATTGTTA	2160
					ACAAGATCCA	2220
					TGAGATTGTA	2280
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ACGGAAACGT	ATCCAACGTA	TCTGTATCAA	AAAGTAGAGG	AAGGTGTATT	AAAACCATAC	2400
					TTATACGATA	2460
					AGATGTATCT	
					GAATAGCCGT	2580
TTAGAAGGAG	AAAACCGTTC	TGGTGATGCA	CATGAGTTCT	CGCTCCCTAT	CGATATAGGA	2640
GAGCTGGATT	ACAATGAAAA	TGCAGGAATA	TGGGTTGGAT	TTAAGATTAC	GGACCCAGAG	2700

	CACHRICCAAA	TCTTGAATTA	GTCGAAGAGG	GACCTTTGTC	AGGAGACGCA	2760
GGATACGCAA	CACILGGAAA	AGAACAACAG	TGGAAGATTC	AAATGACAAG	AAGACGTGAA	2820
TTAGAGCGCT	TGCHAMGHGA	GGCATCGAAA	CARGOGGTAG	ATCGTTTATA	TGCCGATTAT	2880
GAGACAGATA	GAAGATACAT	TGATGTAGAG	AMERICAGATO	TTACTGCGGC	TCAAGATCTG	2940
CAGGATCAAC	AACTGAATCC	TGATGTAGAG	ATTACACATE	BARTACCAGG	GATGAACTAT	3000
ATACAGTCCA	TTCCTTACGT	ATATAACGAA	ATOTTCCCAG	CONTINUE CON	TGATCAGCGA	3060
ACGAAGTTTA	CAGAATTAAC	AGATCGACTC	CAACAAGCG1	AUTOCA ATTOC	AACGCCTGGC	3120
AATGCCATAC	CAAATGGTGA	TTTTCGAAAT	GGGTTAAGTA	MICOLANGE	GGATGAACAA	3180
GTAGAAGTAC	AACAAATCAA	TCATACATCT	GTCCTTGTGA	TTCCAAACTG	macrocca a Ga	3240
GTTTCACAAC	AGTTTACAGT	TCAACCGAAT	CAAAGATATG	TATTACCAGT	TACTOCARGE	3300
AAAGAAGGGG	TAGGAAATGG	ATATGTAAGT	ATTCGTGATG	GTGGAAATCA	ATCAGARACG	3360
CTTACTTTTA	GTGCAAGCGA	TTATGATACA				•
ATCACABABA	CAGTGACATT	•	ACAGATCAAA			3420
ACAGAAGGTA	CGTTCTATAT	AGAAAGTGTA	GAATTGATTG	TAGACGTAGA	G	3471

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:

 (A) LENGTH: 1157 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: YES
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:

 (A) ORGANISM: Bacillus thuringiensis
 (B) STRAIN: kumamotoensis
 (C) INDIVIDUAL ISOLATE: PS50C
- (Vii) IMMEDIATE SOURCE: (B) CLONE: E. coli NM522(pMYC1638), NRRL B-18751
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2: Met Ser Pro Asn Asn Glu Asn Glu Tyr Glu Ile Ile Asp Ala Thr Pro ser Thr Ser Val Ser Ser Asp Ser Asp Arg Tyr Pro Phe Ala Asp Glu Pro Thr Asp Ala Leu Gln Asn Met Asn Tyr Lys Asp Tyr Leu Lys Met 35 Ser Gly Gly Glu Asn Pro Glu Leu Phe Gly Asn Pro Glu Thr Phe Ile ser ser ser Thr Ile Gln Thr Gly Ile Gly Ile Val Gly Arg Ile Leu 65 Gly Ala Leu Gly Val Pro Phe Ala Ser Gln Ile Ala Ser Phe Tyr Ser Phe Ile Val Gly Gln Leu Trp Pro Ser Lys Ser Val Asp Ile Trp Gly 100 Glu Ile Met Glu Arg Val Glu Glu Leu Val Asp Gln Lys Ile Glu Lys 115 Tyr Val Lys Asp Lys Ala Leu Ala Glu Leu Lys Gly Leu Gly Asn Ala 130 Leu Asp Val Tyr Gln Gln ser Leu Glu Asp Trp Leu Glu Asn Arg Asn 145 Asp Ala Arg Thr Arg Ser Val Val Ser Asn Gln Ph Ile Ala Leu Asp 165 170

Leu Asn Phe Val Ser Ser Ile Pr Ser Phe Ala Val Ser Gly His Glu 180 Val Leu Leu Ala Val Tyr Ala Gln Ala Val Asn Leu His Leu Leu 195 Leu Leu Arg Asp Ala Ser Ile Phe Gly Glu Glu Trp Gly Phe Thr Pro 210 220 Gly Glu Ile Ser Arg Phe Tyr Asn Arg Gln Val Gln Leu Thr Ala Glu 225 235 Tyr ser Asp Tyr Cys Val Lys Trp Tyr Lys Ile Gly Leu Asp Lys Leu 255 Lys Gly Thr Thr ser Lys Ser Trp Leu Asn Tyr His Gln Phe Arg Arg 260 270 Glu Met Thr Leu Leu Val Leu Asp Leu Val Ala Leu Phe Pro Asn Tyr 285 Asp Thr His Met Tyr Pro Ile Glu Thr Thr Ala Gln Leu Thr Arg Asp Val Tyr Thr Asp Pro Ile Ala Phe Asn Ile Val Thr Ser Thr Gly Phe 305 Cys Asn Pro Trp Ser Thr His Ser Gly Ile Leu Phe Tyr Glu Val Glu 325 Asn Asn Val Ile Arg Pro Pro His Leu Phe Asp Ile Leu Ser ser Val Glu Ile Asn Thr Ser Arg Gly Gly Ile Thr Leu Asn Asn Asp Ala Tyr 355 365 Ile Asn Tyr Trp Ser Gly His Thr Leu Lys Tyr Arg Arg Thr Ala Asp 370 380 ser Thr Val Thr Tyr Thr Ala Asn Tyr Gly Arg Ile Thr Ser Glu Lys Asn ser Phe Ala Leu Glu Asp Arg Asp Ile Phe Glu Ile Asn Ser Thr 405 Val Ala Asn Leu Ala Asn Tyr Tyr Gln Lys Ala Tyr Gly Val Pro Gly 420 430 ser Trp Phe His Met Val Lys Arg Gly Thr ser ser Thr Thr Ala Tyr Leu Tyr Ser Lys Thr His Thr Ala Leu Gln Gly Cys Thr Gln Val Tyr 450 460 Glu ser ser Asp Glu Ile Pro Leu Asp Arg Thr Val Pro Val Ala Glu 465 475 ser Tyr ser His Arg Leu ser His Ile Thr ser His Ser Phe Ser Lys 495 Asn Gly Ser Ala Tyr Tyr Gly Ser Phe Pro Val Phe Val Trp Thr His Thr ser Ala Asp Leu Asn Asn Thr Ile Tyr ser Asp Lys Ile Thr Gln 515 Ile Pro Ala Val Lys Gly Asp Met Leu Tyr Leu Gly Gly Ser Val Val Gln Gly Pro Gly Phe Thr Gly Gly Asp Ile Leu Lys Arg Thr Asn Pro ser Ile Leu Gly Thr Phe Ala Val Thr Val Asn Gly Ser Leu Ser Gln 575 Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr Asp Phe Glu Phe 580Thr Leu Tyr Leu Gly Asp Thr Ile Glu Lys Asn Arg Phe Asn Lys Thr Met Asp Asn Gly Ala Ser Leu Thr Tyr Glu Thr Phe Lys Phe Ala Ser

Phe Ile Thr Asp Phe Gln Phe Arg Glu Thr Gln Asp Lys Ile Leu Leu 625 630 Ser Het Gly Asp Phe Ser Ser Gly Gln Glu Val Tyr Ile Asp Arg Ile 655 Glu Phe Ile Pro Val Asp Glu Thr Tyr Glu Ala Glu Glu Asp Leu Glu Ala Ala Lys Lys Ala Val Asn Ala Leu Phe Thr Asn Thr Lys Asp Gly 675 Leu Arg Pro Gly Val Thr Asp Tyr Glu Val Asn Gln Ala Ala Asn Leu Val Glu Cys Leu Ser Asp Asp Leu Tyr Pro Asn Glu Lys Arg Leu Leu 705 Phe Asp Ala Val Arg Glu Ala Lys Arg Leu ser Gly Ala Arg Asn Leu 735 Leu Gln Asp Pro Asp Phe Gln Glu Ile Asn Gly Glu Asn Gly Trp Ala Ala Ser Thr Gly Ile Glu Ile Val Glu Gly Asp Ala Val Phe Lys Gly 765 Arg Tyr Leu Arg Leu Pro Gly Ala Arg Glu Ile Asp Thr Glu Thr Tyr Pro Thr Tyr Leu Tyr Gln Lys Val Glu Glu Gly Val Leu Lys Pro Tyr 785 Thr Arg Tyr Arg Leu Arg Gly Phe Val Gly ser ser Gln Gly Leu Glu 815 Ile Tyr Thr Ile Arg His Gln Thr Asn Arg Ile Val Lys Asn Val Pro Asp Asp Leu Leu Pro Asp Val Ser Pro Val Asn Ser Asp Gly Ser Ile 845 Asn Arg Cys Ser Glu Gln Lys Tyr Val Asn Ser Arg Leu Glu Gly Glu 850 Asn Arg Ser Gly Asp Ala His Glu Phe Ser Leu Pro Ile Asp Ile Gly 865 875 Glu Leu Asp Tyr Asn Glu Asn Ala Gly Ile Trp Val Gly Phe Lys Ile Thr Asp Pro Glu Gly Tyr Ala Thr Leu Gly Asn Leu Glu Leu Val Glu 910 Glu Gly Pro Leu Ser Gly Asp Ala Leu Glu Arg Leu Gln Arg Glu Glu 925 Gln Gln Trp Lys Ile Gln Met Thr Arg Arg Arg Glu Glu Thr Asp Arg Arg Tyr Met Ala Ser Lys Gln Ala Val Asp Arg Leu Tyr Ala Asp Tyr 945 Gln Asp Gln Gln Leu Asn Pro Asp Val Glu Ile Thr Asp Leu Thr Ala 965 Ala Gln Asp Leu Ile Gln Ser Ile Pro Tyr Val Tyr Asn Glu Met Phe 980Pro Glu Ile Pro Gly Met Asn Tyr Thr Lys Phe Thr Glu Leu Thr Asp Arg Len Gln Gln Ala Trp Asn Leu Tyr Asp Gln Arg Asn Ala Ile Pro 1010 Asn Gly Asp Phe Arg Asn Gly Leu Ser Asn Trp Asn Ala Thr Pro Gly 1025 Val Glu Val Gln Gln Ile Asn His Thr Ser Val Leu Val Ile Pro Asn 1045 1055 Trp Asp Glu in Val Ser Gln Gln Ph Thr Val Gln Pro Asn Gln Arg 1060 1065

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Tyr Val Leu arg Val Thr Ala Arg Lys Glu Gly Val Gly Asn Gly Tyr 1075 Val Ser lle Arg Asp Gly Gly Asn Gln Ser Glu Thr Leu Thr Phe Ser 1090 1095 Ala ser Asp Tyr Asp Thr Asn Gly Val Tyr Asn Asp Gln Thr Gly Tyr 1105 1115 Ile Thr Lys Thr Val Thr Phe Ile Pro Tyr Thr Asp Gln Met Trp Ile 1130 1135 Ile Val Asp Val Glu 1155

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:

 (A) LENGTH: 1953 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (Vi) ORIGINAL SOURCE:

 (A) ORGANISM: Bacillus thuringiensis
 (B) STRAIN: tolworthi
 (C) INDIVIDUAL ISOLATE: 43F
- (Vii) IMMEDIATE SOURCE:
 (B) CLONE: E. coli ML1-Blue (pM1,98-4), MRRL B-18291
- (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1..1953
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

ATG Met 1	AAT Asn	CCA Pro	AAC Asn	AAT Asn 5	CGA Arg	AGT Ser	GAA Glu	TAT Tyr	GAT Asp 10	ACG Thr	ATA Ile	AAG Lys	GTT Val	ACA Thr 15	CCT PIO	48
AAC Asn	AGT Ser	GAA Glu	TTG Leu 20	CCA Pro	ACT Thr	AAC Asn	CAT His	AAT Asn 25	CAA Gln	TAT Tyr	CCT PIO	TTA Leu	GCT Ala 30	GAC Asp	aat Asn	96
CCA Pro	AAT Asn	TCG Ser 35	ACA Thr	cta Leu	GAA Glu	GAA Glu	TTA Leu 40	AAT Asn	TAT Tyr	AAA Lys	GAA Glu	TTT Phe 45	TTA Leu	AGA Arg	ATG Met	144
ACT Thr	GCA Ala 50	GAC Asp	AAT Asd	TCT Ser	ACG Thr	GAA Glu 55	GTG Val	CTA Leu	GAC Asp	AGC Ser	TCT ser 60	ACA Thi	GTA Val	AAA Lys	GAT Asp	192
GCA Ala 65	GTT Val	GCG GLY	ACA Thr	GGA Gly	ATT Ile 70	TCT Ser	GTT Val	gta Val	GGA Gly	CAG Gln 75	ATT	TTA Leu	ggt gly	GTT Val	GTA Val 80	240.
GGG Gly	GTT Val	CCA Pro	TTT	GCT Ala 85	GGG GGG	GCG Ala	CTC Leu	ACT Thr	TCA ser 90	TTT Phe	TAT Tyr	GAA Gln	TCA Ser	TTT Phe 95	CTT Leu	288
AAC Asn	GCT Ala	ATA Ile	TGG TTD 100	CCA PIO	agt Sei	gat Asp	GCT Ala	GAC Asp 105	CCA Pro	TGG TTP	AAG Lys	GCT Ala	TTT Phe 110	ATG Met	GCA Ala	336
CAA Gln	GTG Val	GAA Glu 115	gta Val	CTG	ATA Ile	gat Asp	AAG Lys 120	AAA Lys	ATA Ile	GAG Glu	GAG Glu	TAT TYT 125	GCT Ala	AAA Lys	AGT Ser	384
AAA Lys	GCT Ala 130	CTT Leu	GCA Ala	GAG Glu	TTA Leu	CAG Gln 135	GGT Gly	CTT Leu	CAA Gln	AAT Asn	AAT ASD 140	TTT	GAA Glu	GAT Asp	TAT Tyr	432

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G	TA al 45	aat Asd	GCG Ala	TTG Leu	gat qaa	TCC Ser 150	TGG TIP	aag Lyb	AAA Lys	GCG Ala	CCT Pro 155	GTA Val	aat abd	TTA Leu	CGA ATG	AGT Ser 160	480
C	GA rg	AGA Arg	AGC Ser	CAA Gln	GAT ASD 165	CGA Arg	ATA Ile	AGA Arg	GAA Glu	CTT Leu 170	TTT	TCT	CAA Gln	GCA Ala	GAA Glu 175	AGC Ser	528
CH	AT Ìs	TTT Phe	CGT ATG	AAT nea 180	TCC	ATG Met	CCG Pro	TCA Ser	TTT Phe 185	GCG Ala	GTT Val	TCC Ser	AAA Lys	TTC Phe 190	GAA Glu	GTT Val	576
C	TG eu	TTT Phe	CTA Leu		ACA Thr	TAT Tyi	GCA Ala	CAA Gln 200	GCT Ala	GCA Ala	TAA IIRA	ACA Thi	CAT His 205	TTA Leu	TTG Leu	CTA Leu	624
T	TA OU	AAA Lys.	GAT Asp	GCT Ala	CAA Gln	gtt Val	TTT Pho 215	GGA Gly	GAA Glu	GAA Glu	TGG TTP	GGA Gly 220	TAT Tyr	TCT Ser	TCA Ser	GAA Glu	.672
						TAT Tyr 230		3.73	727	MITTER.	BAB	والحلم	ACG	CAA	CAA	TAC	720
						AAT Asn										101	768
G	GT Ly	TCA Ser	ACT Thr	TAT TYL 260	GAT Asp	GCA Ala	TGG Trp	GTC Val	AAA Lys 265	TTT Phe	AAC ASD	CGT Arg	TTT Phe	CGC Arg 270	AGA Arg	GAA Glu	816
A	TG at	ACA Thr	TTA Leu 275	ACT Thr	GTA Val	TTA Leu	GAT Asp	CTA Leu 280	ATT Ile	GTA Val	TTA Leu	TTC Phe	CCA Pro 285	TTT Phe	TAT TYT	GAT Asp	864
G!	TT al	CGG Arg 290	TTA Lou	TAC Tyr	TCA Ser	AAA Lys	GGA Gly 295	GTT Val	AAA Lys	ACA Thr	GAA Glu	CTA Leu 300	ACA Thr	AGA Arg	GAC Asp	ATT Ile	912
P	TT he	ACA Thr	gat Asp	CCA Pro	ATT Ile	TTT Phe 310	ACA Thr	CTC	TAA REA	GCT Ala	CTT Leu 315	CAA Gln	GAG Glu	TAT	GGA	Pro 320	960
A	CT hr	TTT Phe	TCG Ser	agt Sei	ATA Ile 325	GAA Glu	AAC Asn	TCT Ser	ATT Tle	CGA Arg 330	AAA Lys	CCT Pro	CAT His	TTA Leu	TTT Phe 335	Asp Asp	1008
T	AT YI	TTG Leu	CGT Arg	GGG Gly 340	ATT Ile	GAA Glu	TTT Phe	CAT	ACG Thr 345	CGT AIG	CTT Leu	CGA Arg	Pro	GGT Gly 350	TAC Tyr	TCT Ser	1056
G	GG ly	AAA Lys	GAT Asp 355	TCT Ser	TTC Phe	aat asn	TAT Tyr	TGG Trp 360	TCT Ser	GGT Gly	AAT ASII	TAT Tyr	GTA Val 365	GAA Glu			1104
C.	CT TO	AGT Ser	ATA Ile	gga Gly	TCT Ser	TAA NBA	GAT ASP 375	ACA Thr	ATC Ile	ACT	TCC	CCA Pro 380	TTT	TAT	GGA Gly	Asp	1152
A L	AA VB	TCT Ser	ATT Ile	GAA Glu	CCT Pro	ATA Ile 390	CAA Gln	AAG Lys	CTA	AGC Ser	TTT Phe 395	gat Asp	GLY	CAA Gln	Lys	Val 400	1200
T	ŊΕ	Arg	THE	146	405					410			٠.		415		1248
		_		420												CAA Gln	1296
L	ys	Asn	435	The	ser	ACA Thr	GIII	440	-1-				445	•			1344
T	Ϋ́	450	GTĀ	ALA	GIII	GAT Asp	455	110	ern P			460		٠			1392
G						AAA Lys 470	GCA Ala	m> m	300	CAM	CAC	COL	AAT	TAC	GCA	GAA	1440

TGT Cyb	TTC Phe	TTA Leu	ATG Met	CAG Gln 485	GAC Asp	CGT Arg	CGT Arg	GGA Gly	ACA Thr 490	ATT Il	CCA Pro	TTT	TTT Phe	ACT Thr 495	TGG Trp		1488
ACA Thr	CAT His	AGA Arg	AGT Ser 500	GTA Val	GAC Asp	TTT Phe	TTT Phe	AAT Asn 505	ACA The	ATT Ile	gat Asp	GCT Ala	GAA Glu 510	AAA Lys	ATT Ile		1536
ACT Thr	CAA Gln	CTT Leu 515	CCA Pro	gta Val	val Val	AAA Lys	GCA Ala 520	TAT Tyr	GCC Ala	TTG Lou	TCT Ser	TCA Ser 525	Gly	GCT Ala	TCC Ser		1584
ATT Ile	ATT Ile 530	GAA Glu	Gly	CCA Pro	GGA Gly	TTC Pho 535	ACA Thr	GGA Gly	GGA Gly	AAT Asn	TTA Leu 540	CTA	TTC Phe	CTA Leu	AAA Lys	:	1632
GAA Glu 545	TCT Ser	AGT Sei	AAT Asn	TCA Sei	ATT Ile 550	GCT Ala	AAA Lys	TTT Phe	AAA Lyb	GTT Val 555	ACC Thr	TTA Leu	ART	TCA Ser	GCA Ala 560	:	1680
ecc Ala	TTG Lou	TTA Lou	CAA Gln	CGA Arg 565	TAT Tyr	CGC	GTA Val	AGA Arg	ATA 110 570	Arg	TAT Tyr	GCT Ala	TCA Sei	ACC Thr 575	ACT	3	1728
AAC Asn	CTA Leu	CGA A rg	CTT Leu 580	TTC Pho	GTG Val	CAA Gln	AAT ABD	TCA Ser 585	AAC Asn	AAT Asn	GAT Asp	TTT Phe	CTT Leu 590	GTC Val	ATC Ile	;	1776
TAC Tyr	ATT Ile	AAT Aan 595	AAA Lys	ACT Thr	ATG Met	AAT Asd	ATA Ile 600	GAT Asp	GGT Gly	gat Asp	TTA Leu	ACA Thr 605	TAT Tyr	CAA Gln	ACA Thr	1	L824
TTT Phe	GAT ABP 610	TTC Phe	GCA Ala	ACT Thr	AGT Ser	AAT Asn 615	TCT Set	TAA nea	ATG Met	GGA Gly	TTC Pho 620	TCT Ser	GGT Gly	GAT Asp	ACA Thr	:	1872
AAT ABD 625	GAC Asp	TTT Phe	ATA Ile	ATA Ile	GGA Gly 630	GCA Ala	GAA Glu	TCT Ser	TTC Phe	GTT Val 635	TCT	AAT ASD	GAA Glu	AAA Lys	ATC Ile 640	1	L920
TAT Tyr	ATA Ile	GAT Asp	AAG Lys	ATA Ile 645	GAA Glu	TTT Phe	ATC 11e	CCA Pro	GTA Val 650	CAA Gln						1	1953

- (2) INFORMATION FOR SEQ ID NO:4:
 - (i) SEQUENCE CHARACTERISTICS:

 (A) LENGTH: 651 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (iii) HYPOTHETICAL: YES
 - (iv) ANTI-SENSE: NO
 - (vi) ORIGINAL SOURCE:
 (A) ORGANISM: Bacillus thuringiensis
 (B) STRAIN: tolworthi
 (C) INDIVIDUAL ISOLATE: 43F
 - (vii) IMMEDIATE SOURCE:
 (B) CLONE: E. Coli ML1-Blue (pM1,98-4), NRRL B-18291
 - (ix) FEATURE:
 (A) NAME/KEY: Protein
 (B) LOCATION: 1..651
 - (Mi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Asn Pro Asn Asn Arg Ser Glu Tyr Asp Thr Ile Lys Val Thr Pro 15 As n Ser Glu Leu Pro Thr As nHis As n Gln Tyr Pro Leu Ala Asp As n 20° Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met 35

Thr ala Asp Asn Ser Thr Glu Val Leu Asp Ser Ser Thr Val Lys Asp 50 Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val 65 Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu Asn Ala Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr Val Asn Ala Leu Asp Ser Trp Lys Lys Ala Pro Val Asn Leu Arg Ser 145 150 150 Arg Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu ser 165 170 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val Leu Phe Leu Pro Thr Tyr Ala Gin Ala Ala Asn Thr His Leu Leu Leu 195 205 Leu Lys Asp Ala Gln Val Pha Gly Glu Glu Trp Gly Tyr ser ser Glu Asp Ile Ala Glu Phe Tyr Glu Arg Glu Leu Lys Leu Thr Glu Glu Tyr 225 230 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Ser Leu Arg 255 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 285 Val Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 295 Phe Thr Asp Pro Ile Phe Thr Leu Asn Ala Leu Gln Glu Tyr Gly Pro 320 Thr Phe ser ser Ile Glu Asn ser Ile Arg Lys Pro His Leu Phe Asp 335 335 Tyr Leu Arg Gly Ile Glu Phe His Thr Arg Leu Arg Pro Gly Tyr Ser Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg Pro Ser Ile Gly Ser Asn Asp Thr Ile Thr Ser Pro Phe Tyr Gly Asp 370 Lys ser lle Glu Pro Ile Gln Lys Len ser Phe Asp Gly Gln Lys Val Tyr Arg Thr Ile Ala Asn Thr Asp Ile $\frac{1}{410}$ Ala Phe Pro Asp Gly Lys $\frac{1}{415}$ Ile Tyr Phe Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln 430 Lys Ash Glu Thr ser Thr Gln Thr Tyr Asp Ser Lys Arg Tyr Ash Gly Tyr Leu Gly Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr 450 455 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu 480 Cys Phe Leu Met Gin Asp Arg Arg ly Thr Ile Pro Phe Phe Thr Trp

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Thr His Arg ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile 500 510 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu ser ser Gly Ala ser Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys 530 Glu ser ser Asn ser Ile Ala Lys Phe Lys Val Thr Leu Asn ser Ala 545 555 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr 575 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile 585 Tyr Ile Asn Lys Thr Met Asn Ile Asp Gly Asp Leu Thr Tyr Gln Thr Phe Asp Phe Ala Thr ser Asn ser Asn Met Gly Phe Ser Gly Asp Thr 610 620 Asn Asp Phe Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile 625 635 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln 650

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:

 (A) LENGTH: 1425 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 (A) ORGANISM: BACILLUS THURINGIENSIS
 (C) INDIVIDUAL ISOLATE: PS86A1
- (Vii) IMMEDIATE SOURCE:
 (B) CLONE: E. COli NM522(pMYC2320) NRRL B-18769
- (ix) FEATURE:
 (A) NAME/KEY: mat peptide
 (B) LOCATION: 1..1425
- (Ri) SEQUENCE DESCRIPTION: SEQ ID NO:1: 60 ARTTCTARTA AGARATATGG TCCTGGTGAT ATGACTARTG GARATCARTT TATTATTTCA 120 AAACAAGAAT GGGCTACGAT TGGAGCATAT ATTCAGACTG GATTAGGTTT ACCAGTAAAT 180 GAACAACAAT TAAGAACACA TGTTAATTTA AGTCAGGATA TATCAATACC TAGTGATTTT 240 TCTCAATTAT ATGATGTTTA TTGTTCTGAT AAAACTTCAG CAGAATGGTG GAATAAAAAT 300 360 TTATATCCTT TAATTATTAA ATCTGCTAAT GATATTGCTT CATATGGTTT TAAAGTTGCT GGTGATCCTT CTATTAAGAA AGATGGATAT TTTAAAAAAT TGCAAGATGA ATTAGATAAT 420 ATTGTTGATA ATAATTCCGA TGATGATGCA ATAGCTAAAG CTATTAAAGA TTTTAAAGCG 480 CGATGTGGTA TTTTAATTAA AGAAGCTAAA CAATATGAAG AAGCTGCAAA AAATATTGTA 540 ACATCTITAG ATCAATTITT ACATGGTGAT CAGAAAAAT TAGAAGGTGT TATCAATATT 600 CARALACGIT TARAAGAAGI TCAAACAGCI CITAATCAAG CCCATGGGGA AAGTAGICCA 660 GCTCATAAAG AGTTATTAGA AALAGTAAAA AATTTAAAAA CAACATTAGA AAGGACTATT 720 Anagotgaac aagatttaga gaaaaaagta gaatatagtt ttotattagg accattgtta 780

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CGATTIGTTG TTTATGAAAT	TCTTGAAAAT	ACTGCTGTTC	AGCATATAAA	AAATCAAATT	840
GATGAGATAA AGAAACAATT					900
ATAGGAATGT TAAATAGTAT	MARINACACAM.	አመተረስሞሽስጥም	TATATAGTCA	AGGACAAGAA	960
ATAGGAATGT TAAATAGTAT	THATALAGAI	Water and a series of the seri		001110101	1020
GCAATTAAAG TTTTCCAAAA	GTTACAAGGT	ATTTGGGCTA	CTATTGGAGC	TCAAATAGAA	
AATCTTAGAA CAACGTCCTT	ACAAGAAGTT	CARGATTCTG	ATGATGCTGA	TGAGATACAA	1080
ATTGAACTTG AGGACGCTTC	•				1140
	_			TATATCAGAT	1200
	TAATAGTAGA				
TCATGTAATT GTTCAACAAC	BARTATGACA	TCARATCART	ACAGTAATCC	AACAACAAAT	1260
ATGACATCAA ATCAATATAT	GATTTCACAT	GLATATACAA	GTTTACCAAA	TAATTTTATG	1320
				GATATATTGG	1380
TTATCARGAA ATAGTAATIT			25,122,22	CHEATING & CO.	
TATAATAATT CGGATTGGTA	TAATAATTCG	GATTGGTATA	ATAAT	•	1425

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:

 (A) LENGTH: 475 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: YES
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 (A) ORGANISM: BACILLUS THURINGIENSIS
 (C) INDIVIDUAL ISOLATE: PS86Al
- (VII) IMMEDIATE SOURCE: (B) CLONE: E. coli NM522(pMYC2320) NRRL B-18769
- (ix) FEATURE:
 (A) NAME/REY: Protein
 (B) LOCATION: 1..475

(xi) SEQUENCE DESCRIPTION: SEQ ID No:2: Met Ile Ile Asp Ser Lys Thr Thr Leu Pro Arg His Ser Leu Ile His Thr Ile Lys Leu Asn Ser Asn Lys Lys Tyr Gly Pro Gly Asp Met Thr 20 25 Asn Gly Asn Gln Phe Ile Ile Ser Lys Gln Glu Trp Ala Thr Ile Gly $\frac{45}{45}$ Ala Tyr Ile Gln Thr Gly Leu Gly Leu Pro Val Asn Glu Gln Gln Leu 50 Arg Thr His Val Asn Lau Ser Gln Asp Ile Ser Ile Pro Ser Asp Phe 80 ser Gln Leu Tyr Asp Val Tyr Cys Ser Asp Lys Thr Ser Ala Glu Trp $_{95}^{85}$ Trp Asn Lys Asn Leu Tyr Pro Leu Ile Ile Lys Ser Ala Asn Asp Ile 105 Ala Ser Tyr Gly Phe Lys Val Ala Gly Asp Pro Ser Ile Lys Lys Asp 125 Gly Tyr Phe Lys Lys Leu Gln Asp Glu Leu Asp Asn Ile Val Asp Asn 130 140 Asn Ser Asp Asp Asp Ala Ile Ala Lys Ala Ile Lys Asp Phe Lys Ala 145 Arg Cys Gly Ile Leu Ile Lys Glu Ala Lys Gln Tyr Glu Glu Ala Ala 175 Lys Asn Ile Val Thr ser Leu Asp Gln Phe Leu His Gly Asp Gln Lys

185 180 Lys Leu Glu Gly Val Il Asn Ile Gln Lys Arg Leu Lys Glu Val Gln 195 205 Thr Ala Leu Asn Gln Ala His Gly Glu Ser Ser Pro Ala His Lys Glu 210 220 Leu Leu Glu Lys Val Lys Asn Leu Lys Thr Thr Leu Glu Arg Thr Ile 225 230 230 Lys Ala Glu Glu Asp Leu Glu Lys Lys Val Glu Tyr Ser Phs Leu Leu 255 Gly Pro Leu Leu Gly Phe Val Val Tyr Glu Ile Leu Glu Asn Thr Ala Val Gln His Ile Lys Asn Gln Ile Asp Glu Ile Lys Lys Gln Leu Asp ser Ala Gln His Asp Leu Asp Arg Asp Val Lys Ile Ile Gly Met Leu 295 Asn Ser Ile Asn Thr Asp Ile Asp Asn Leu Tyr Ser Gln Gly Gln Glu 305 Ala Ile Lys Val Phe Gln Lys Leu Gln Gly Ile Trp Ala Thr Ile Gly 335 Ala Gln Ile Glu Asn Leu Arg Thr Thr Ser Leu Gln Glu Val Gln Asp ser Asp Asp Ala Asp Glu Ile Glu Ile Glu Leu Glu Asp Ala Ser Asp Ala Trp Leu Val Val Ala Gln Glu Ala Arg Asp Phe Thr Leu Asn Ala Tyr ser Thr Asn ser Arg Gln Asn Leu Pro Ile Asn Val Ile Ser Asp 385 ser Cys Asn Cys Ser Thr Thr Asn Met Thr Ser Asn Gln Tyr Ser Asn 415 Pro Thr Thr Asn Met Thr Ser Asn Gln Tyr Met Ile Ser His Glu Tyr 420Thr Ser Leu Pro Asn Asn Phe Met Leu Ser Arg Asn Ser Asn Leu Glu 435 Tyr Lys Cys Pro Glu Asn Asn Phe Met Ile Tyr Trp Tyr Asn Asn Ser 450 460 Asp Trp Tyr Asn Asn Ser Asp Trp Tyr Asn Asn 465

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 53 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
 (A) ORGANISM: Bacillus thuringiensis
 (B) STRAIN: PS86A1
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

ATGATTGATT CTARARCARC ATTACCARGA CATTCHTTAR TWCATACHAT WAR

<u>Claims</u>

1	1. A method for controlling lepidopteran insect pests which comprises contacting said
	is cost pasts with an insect-controlling effective amount of a Bacillus thuringiensis isolate selected
2	from the group consisting of Bt. PS43F, Bt. PS50C and Bt. PS86A1, and variants thereof.
3	non the group washing of 2
1	2. The method, according to claim 1, wherein said Bacillus thuringiensis is Rt. PS43F.
ľ	3. The method, according to claim 1, wherein said Bacillus thuringiensis is B.t. PS50C.
1	4. The method, according to claim 1, wherein said Bacillus thuringiensis is B.t. PS86A1.
1	5. The method, according to claim 1, wherein said insect pest is the diamondback moth
2	(Piuteila xylostella).
1	6. The method, according to claim 1, which comprises applying an insecticidal
2	composition comprising Bacillus thuringiensis to plants or soil.
1	7. The method, according to claim 6, wherein said insecticidal composition is a liquid.
1	8. The method, according to claim 6, wherein said insecticidal composition is in granular
2	form.
1	9. The method, according to claim 6, wherein said insecticidal composition is applied
2	when corn or soybean seed is planted.
1	10. The method, according to claim 1, wherein said Bacillus thuringiensis are treated to
	prolong their pesticidal activity in the environment of a target pest.
2	
1	11. A method for controlling lepidopteran pests which comprises exposing said pests to
1	a start transformed by a sene obtainable from a Bacillus thuringiensis isolate selected from the
2	group consisting of Rt. PS43F, Rt. PS50C and Rt. PS86A1, and variants thereof, wherein said
3	gene encodes a toxin active against lepidopteran pests.
4	
	12. The method, according to claim 11, wherein said gene comprises the DNA of SEQ
1	ID NO. 1 or a portion thereof which encodes a lepidopteran-active toxin.
2	ID MO. I of a hormy support

1	13. The method, according to claim 11, wherein said gene comprises the 2-th comprises
2	ID NO. 3 or a portion thereof which encodes a lepidopteran-active toxin.
1	14. The method, according to claim 11, wherein said gene comprises the DNA of SEQ
2	ID NO. 5 or a portion thereof which encodes a lepidopteran-active toxin.
1	15. A method for controlling lepidopteran insects which comprises administering to said
2	insects or to the environment of said insects a microorganism transformed by a gene obtainable
3	from a Bacillus thuringiensis isolate selected from the group consisting of B.t. PS43F, B.t. PS50C
4	and R.t. PS86A1, and variants thereof, wherein said gene encodes a toxin active against
5	lepidopteran pests.
1	16. The method, according to claim 15, wherein said gene comprises the DNA of SEQ
2	ID NO. 1 or a portion thereof which encodes a lepidopteran-active toxin.
1	17. The method, according to claim 15, wherein said gene comprises the DNA of SEQ
2	ID NO. 3 or a portion thereof which encodes a lepidopteran-active toxin.
1	18. The method, according to claim 15, wherein said gene comprises the DNA of SEQ
2	ID NO. 5 or a portion thereof which encodes a lepidopteran-active toxin.
1	19. The method, according to claim 15, wherein said microorganism is a Pseudomonas.
1	20. The method, according to claim 15, wherein said transformed microorganism is
2	treated to prolong its pesticidal activity in the environment of a target pest.

INTERNATIONAL SEARCH REPORT

International Application No PCT/US 92/07697

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P,X	EP, A2, 0498537 (MYCOGEN CORPORATION	,							
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x	EP, A1, 0337604 (MYCOGEN CORPORATION)	1-20						
^	l 18 October 1989.		i i						
}	see the claims; page 2, lines 30	-37							
			1						
		A1 \	1-20						
x	US, A, 4849217 (GEORGE G. SOARES ET	AL.)							
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Form PCT	ISA/210 (second sheet) (January 1985)								

L DOCL	MENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET) Clistian of Document, with indication, where appropriate, of the relevant passages	Relevant to Cislm No
resides.	US. A. 4996155 (AUGUST J. SICK ET AL.)	1-20
	26 February 1991, see the whole document	
	·	
,х	EP, A2, 0475656 (MYCOGEN CORPORATION) 18 March 1992, see the claims	1-20
	see the Claims	
,x	EP, A2, 0500311 (MYCOGEN CORPORATION) 26 August 1992, see the claims	1-20
	366 016 214 VIII	
	EP, A2, 0401979 (MYCOGEN CORPORATION)	1-20
	12 December 1990, see the claims	
-	EP, A2, 0303379 (MYCOGEN CORPORATION) 15 February 1989, see the claims	1-20
	266 PHG #14149	
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ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO.PCT/US 92/07697

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This annex fixts the patent family members relating to the satent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on

The European Patent office is in so way fiable for these particulars which are merely given for the purpose of information.

Patent document cited in search report	Publication data	Patent fo mambe	Publication data	
EP-A2- 0498537	12/08/92	NONE		
EP-A1- 0337604	18/10/89	JP-A- US-A- US-A- US-A- US-A-	2005874 4996155 5064648 5104974 5133962	10/01/90 26/02/91 12/11/91 14/04/92 28/07/92
US-A- 4849217	18/07/89	NONE		
US-A- 4996155	26/02/91	EP-A- JP-A- US-A- US-A- US-A-	0337604 2005874 5064648 5104974 5133962	18/10/89 10/01/90 12/11/91 14/04/92 28/07/92
EP-A2- 0475656	18/03/92	AU-D- US-A- US-A- US-A- US-A- US-A-	8378391 5064648 5093119 5098705 5100665 5106620	19/03/92 12/11/91 03/03/92 24/03/92 31/03/92 21/04/92
EP-A2- 0500311	26/08/92	NONE		
EP-A2- 0401979	12/12/90	CA-A- JP-A-	2015951 3081210	18/11/90 05/04/91
EP-A2- 0303379	15/02/89	JP-A- US-A-	1050806 4910016	27/02/89 20/03/90